

THE NEW ARMENIAN MEDICAL JOURNAL

Volume19 (2025), Issue 1, p. 31-37

(1) 95

DOI: https://doi.org/10.56936/18290825-1.v19.2025-31 THE EFFECT OF THE MEDICINAL COMPOSITION "EFLORNITHINE-ARMENICUM" ON THE PROGRESSION OF THE INFLAMMATORY PROCESS IN AN EXPERIMENTALLY INDUCED AEROBIC WOUND

GHAZARYAN H.V.

Arpimed Pharmaceutical Company LLC, Abovyan, Armenia Received 22.06.2024; Accepted for printing 11.02.2025

ABSTRACT

Wound infection remains one of the most serious current challenges in modern medicine. Significant challenges in the symptomatic and pathogenetic therapy of wound infections arise due to the known symbiosis between pathogenic and opportunistic bacteria and certain pathogenic and opportunistic fungi. The mixed bacterial-fungal microflora persisting during a wound infection is often described as a biofilm. Notably, the addition of a fungal infection significantly worsens the wound healing process: on one hand, fungi that persist in the host's wound are inherently toxic; on the other, their association with bacteria often enhances the pathogenic potential of the bacteria.

The therapeutic efficacy of the medicinal composition Eflornithine-Armenicum was studied using an experimentally induced aerobic wound model. This medicinal composition was developed at the Research Center of the Yerevan State Medical University in collaboration with Arpimed LLC.

A wide range of morphological, morphometric, cytological, bacteriostatic, and immunomorphological studies were conducted. It was found that three applications of the composition to the wound surface on the skin of experimental rats led to an early activation of reparative and proliferative processes, ultimately resulting in complete restoration of the integrity of the damaged wound tissues through substitution.

The therapeutic effectiveness of the tested medicinal composition is, on one hand, due to the pronounced antibacterial activity of Eflornithine, which facilitated the early self-cleansing of the wound from opportunistic and pathogenic microorganisms persisting in situ. On the other hand, the effectiveness is attributed to the strong anti-inflammatory activity of Armenicum paste, thanks to the presence of ionized iodine in its composition.

Based on our studies, we believe there are broad prospects for further preclinical and clinical research on Eflornithine-Armenicum as an effective therapeutic agent for the pathogenetic treatment of wound inflammation.

Based on our comprehensive studies, we conclude that the medicinal composition Eflornithine-Armenicum, which we developed, should be considered an effective therapeutic agent in the treatment of aerobic wounds. This is particularly important, as both components of the composition have long been approved by prestigious pharmaceutical regulatory bodies as medicinal products with confirmed effectiveness and safety.

Keywords: aerobic wound, wound infections, medicinal composition "Eflornithine-Armenicum, treatment.

CITE THIS ARTICLE AS:

Ghazaryan H.V. (2025). The effect of the medicinal composition "Eflornithine-Armenicum" on the progression of the inflammatory process in an experimentally induced aerobic wound. The New Armenian Medical Journal, vol.19(1), 31-37; DOI: https://doi.org/10.56936/18290825-1.v19.2025-31

ADDRESS FOR CORRESPONDENCE: Hovhannes V. Ghazaryan Arpimed Pharmaceutical Company LLC 2nd Micro-District, 19 b., Abovyan 2204, Kotayk Marz, Armenia Tel.: (374 41) 020510 E-mail: hovhannes.ghazaryan@arpimed.com

INTRODUCTION

Wound infections remain a significant and serious problem in modern medicine. The situation is further complicated by the changing nature of pathological processes and the resistance of opportunistic and pathogenic microorganisms in wounds to many antibacterial drugs, especially antibiotics [*Pfaller M, 2012, Kapoor G et al., 2017; Reygaert W, 2018; Berman J, Krysan D, 2020*].

Significant challenges in symptomatic and pathogenetic therapy during wound infection arise from the known symbiosis of pathogenic and opportunistic bacteria, as well as certain fungi. This mixed bacterial-fungal microflora in wound infections is often described as a biofilm [James G et al., 2008, Dowd S et al., 2011; Clinton A, Carter T, 2015; Percival S et al., 2015, Michael AJ, 2018, Rocha R, Wilson R, 2018]. It is particularly noteworthy that the addition of a fungal infection significantly worsens the progression of the wound healing process. On one hand, the fungi persisting in the host wound are inherently toxic; on the other, when in association with bacteria, they often enhance the pathogenic potential of the bacterial presence. Unfortunately, current antifungal therapies are not always effective [Becker W, 1991, Pruskowski K et al., 2021].

The current situation regarding daily diet choices is concerning, as various supplements, including hormonal ones, are commonly used to increase the meat mass of poultry, fish, cattle and small ruminants [*Fritsche S, Steinhart H, 1999, Saha S, Pathak N, 2021*].

As a result of the factors mentioned above, the search for effective agents, and particularly the development of medicinal compositions with a broad, multipotent spectrum of action, represents a promising and relevant scientific and practical approach in modern medicine.

At the research center of Yerevan State Medical University named after M. Herats and Arpimed, LLC (Abovyan, Armenia), we have developed and successfully completed preclinical testing of the medicinal composition "Eflornithine (DFMO)-Armenicum" [*Ghazaryan H, Hovhannisyan A, 2022*].

The selection of agents for the medicinal composition was determined by the following consideration. It is well established that DFMO exhibits a strong inhibitory effect, suppressing the synthesis of aliphatic polyamines at the earliest stages of their enzymatic transformation, specifically, the conversion of ornithine to putrescine [*Meyskens F*, *Gerner E*, 1999; *Gerner E*, *Meyskens F*, 2009].

It is also worth noting that in recent years, highly informative data has emerged indicating that the vital activity and persistence of many resident pathogenic and opportunistic bacteria, viruses, and fungi within the host body are largely maintained by mechanisms that are specifically polyamine-dependent [Wallace H, Fraser A, 2004; Shah P, Swiatlo E, 2008; Wallace H, 2009; Valdés-Santiago L et al., 2012; Valdés-Santiago L, Ruiz-Herrera J, 2014; Bae D et al., 2018; Berman J, Krysan D, 2020]. In this context, there have been rare attempts to use DFMO to inhibit the synthesis of aliphatic polyamines not only in somatic cells but also to suppress their synthesis within microbial cells [Wallace H, Fraser A, 2004; Wallace H, 2009; Bae D et al., 2018; Berman J, Krysan D, 2020].

The second component of the medicinal composition, *Armenicum* paste, has a notably pronounced anti-inflammatory and partially bacteriostatic spectrum of action [*Zilfyan A et al., 2016*].

MATERIAL AND METHODS

The investigation involved 180 male Wistar rats, in which an aerobic wound model was induced according to the method proposed by Hovhannisyan S.S. et al., for which a patent has been granted [*Hovhannisyan S et al.*, 1987].

The animals in both the experimental and control groups were divided into three subgroups, which were removed from the experiment on the third, fifth, and ninth days of the investigation. The control group received only *Armenicum* paste, applied to the wound surface three times at 4-hour intervals, at a dose of 5.1 mg/kg. In addition to *Armenicum* paste, the experimental group also received the medicinal composition Eflornithine (DFMO)-*Armenicum*, applied to the wound surface three times at the same intervals. A single dose of *Armenicum* paste was 5.1 mg/kg, while a single dose of Eflornithine was 460 mg/kg.

The study employed conventional morphological methods, including staining with azure-II eosin and hematoxylin-eosin. Bacteriological methods were also used, incorporating staining with azure-II eosin and fluorochromizing with acridine orange. An immunomorphological method was utilized to detect fibronectin in tissues, specifically the indirect Coons method with rabbit anti-fibronectin serum (Sigma, USA) and FITC-labeled anti-rabbit IgG serum (Sigma, USA). The preparations were examined under a trinocular light microscope (Micros, Austria) and a trinocular fluorescence microscope (Boeco, Germany).

Statistical analysis was performed using the SPSS program, 13 ANOVA version, using Student's t-criteria.

RESULTS AND DISCUSSION

The study results indicated that local treatment of skin wounds in the experimental rats led to a marked activation of local reparative-proliferative processes. In contrast to the control group, which received only *Armenicum* paste, the experimental group showed activation of reparative-proliferative processes as early as the third day of the investigation. In the control group, where only *Armenicum* paste was applied to the wound surface, processes aimed at restoring defect integrity were observed only on the fifth and ninth days of the regional inflammatory process.

By the third day of the experiment, reparative processes were evident through the focal development of granulation tissue, which by the fifth day had become more widespread and showed increased differentiation (Fig. 1 a, b, c, d).

A clear trend toward differentiation of granu-

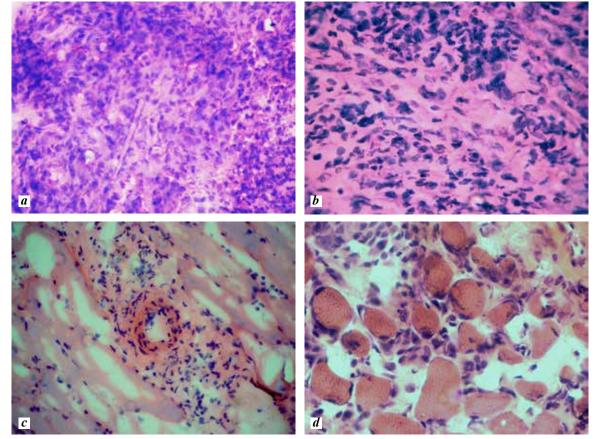


Figure 1. Structural changes in the soft tissues of the wound in animals from the experimental group on the 5th day of investigation.

(a). Further differentiation of granulation tissue transitioning to loose connective tissue in the superficial areas of the wound. Hematoxylin-eosin staining. Oc. 15, Ob. 60.

(b). Against a background of moderate edema, signs of granulation tissue growth with a tendency toward an organized structure of newly formed collagen fibers are observed. Stained with azure II-eosin. Oc. 15, Ob. 60.

(c). Productive subacute vasculitis with perivascular myocytolysis in the underlying muscle tissue. Stained with hematoxylin - eosin. Oc. 10, Ob. 10.

(d). Edema and cellular infiltration of the intermuscular tissue, with dystrophic changes observed in a distinct group of myocytes. Stained with hematoxylin and eosin. Oc 10, Ob. 60.

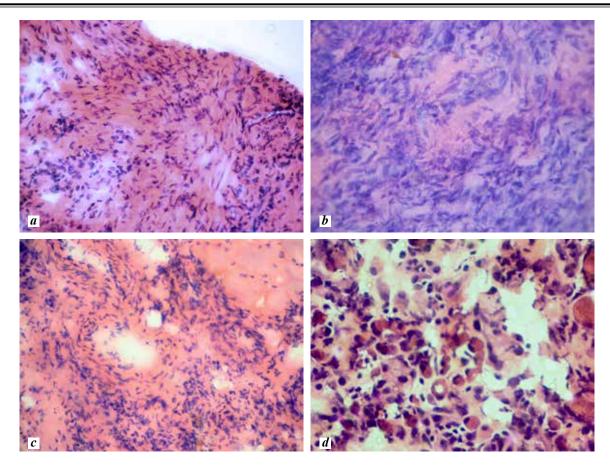


Figure 2. Structural changes in the soft tissues of the wound in control group animals. Stained with hematoxylin - eosin. 5th day of investigation.

(a). Dystrophic changes in inflammatory and connective tissue cells in the superficial areas of the wound, with early focal signs of granulation tissue revitalization in these regions. Oc. 15, Ob. 20.

(b). Poorly differentiated strands of granulation tissue with randomly oriented collagen fibers. Oc. 15, Ob. 20.

(c). Moderate perivascular infiltration of inflammatory cells, with focal activation of fibroblastic cells. Oc. 15, Ob. 10.

(d). Single muscle cells are visible in cross-section. The intermuscular spaces are compressed and infiltrated with inflammatory cells. Oc. 15, Ob. 60.

lation tissue was observed. In the control group animals, granulation tissue remained detectable even at the later stages of the investigation. Additionally, catabolic processes were observed more frequently in the control group, marked by areas of necrobiosis and death of connective elements, including fibroblastic and angiomatous cells, as well as myocytes (Fig. 2 a, b, c, d).

At relatively late stages of the regional inflammatory process (the 9th day of investigation), the wound induced in rats healed through substitution, meaning the integrity of the soft tissue covering the wound defect was restored by secondary intention and connective tissue growth (Fig. 3 a, b).

What are the possible mechanisms underlying the beneficial effect of the medicinal composition

"Eflornithine-Armenicum" on the recovery process in an induced aerobic wound?

Primarily, the positive effect of local application of this medicinal composition can be attributed to the direct action of its components on the bacterial landscape of the wound. Notably, the composition exhibited a pronounced bactericidal effect, simultaneously targeting both pathogenic and opportunistic microflora persisting in the wound. This observation is supported by our bacterioscopic analysis of the wound exudate microflora. As mentioned earlier, many opportunistic and pathogenic microorganisms, including bacteria residing in the wound exudate and soft tissues, particularly in areas of tissue destruction, require aliphatic polyamines to support their reproduction and vital functions.

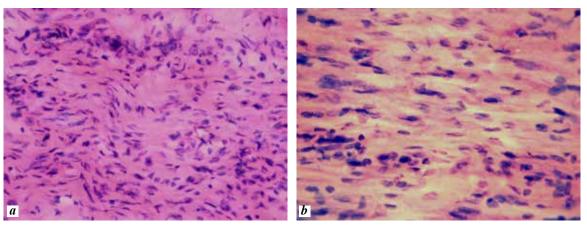


Figure 3. Structural changes in the soft tissues of the wound in experimental group animals. 9th day of investigation. (a). Differentiated connective tissue is present in the superficial parts of the wound. Collagen fibers acquire a linear, ordered orientation, with mature fibrocytes beginning to predominate among the fibroblastic cells. Stained with hematoxylin - eosin. Oc 15, Ob. 20.

(b). Further differentiation of granulation tissue in the deep parts of the wound. Stained with hematoxylin and eosin. Oc. 15, Ob. 10.

It is particularly noteworthy that the pronounced bactericidal effect observed was primarily due to DFMO in the medicinal composition, which facilitated rapid cleansing of the wound from persisting microorganisms. This effect was confirmed by our cytological and bacterioscopic studies, using azure-II eosin and acridine orange staining (Fig. 4 a, b).

We also observed that, with the cleansing of the wound from microorganisms, the structural and functional characteristics of wound exudate cells, macrophages, leukocytes, and lymphocytes, became significantly normalized. Concurrently, complete phagocytosis was activated in structurally intact macrophages within the wound exudate. This facilitated early activation of reparative and proliferative processes in the soft tissue surrounding the wound, ultimately leading to full healing by substitution, wherein the entire length of the damaged tissues was replaced by loose connective tissue.

Additionally, our immunomorphological studies revealed that the presence of effornithine in the medicinal composition promoted pronounced syn-

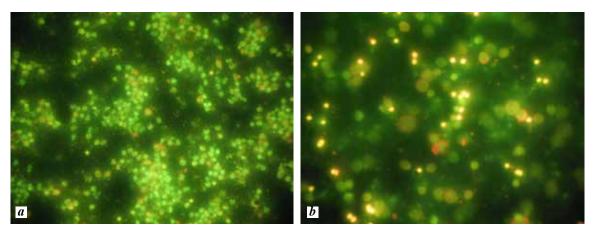


Figure 4. Structural changes in immunocompetent cells of wound exudate in experimental group animals. Stained with acridine orange. Oc. 15, Ob. 20

(a). Single macrophages and a moderate number of lympho-leukocyte cells are present, with changes in their tinctorial properties: green fluorescence shifts to orange-red. Extracellularly oriented single green and orange-red granules are also observed. 3rd day of investigation.

(b). Structurally preserved immunocompetent cells with green fluorescence dominate in the exudate. 5th day of investigation.

The New Armenian Medical Journal, Vol. 19 (2025), Is. 1, p. 31-37

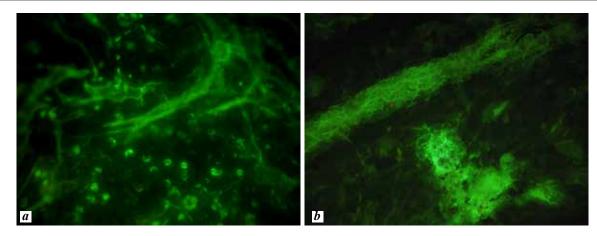
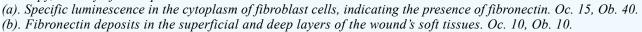


Figure 5. Presence of fibronectin in the soft tissues of the wound in experimental group animals. Luminescent microscopy, 3rd day of the experiment.



thesis of fibronectin by fibroblasts in the wound's soft tissues, even at the early stages of the inflammatory process (Fig. 5 a, b).

Fibronectin is a well-known potent activator of fibroblast cell proliferation, which, in turn, initiates the production of collagen protein structures during the wound inflammatory process [*Grinnell F et al., 1981; Lenselink E, 2015; Gimeno-LLuch I et al., 2022*]

CONCLUSION

Based on our comprehensive studies, we conclude that the medicinal composition "*Eflornithine-Armenicum*" should be considered an effective therapeutic agent for treating aerobic wounds. This is particularly significant, as both components have been approved by prestigious pharmaceutical regulatory bodies, affirming their effectiveness and safety as medicinal products.

REFERENCES

- Bae DH, Lane D, Jansson PJ, Richardson DR (2018). The old and new biochemistry of polyamines. Biochimica et Biophysica Acta (BBA)
 General Subjects. S0304416518301661. DOI: 10.1016/j.bbagen.2018.06.004
- Becker WK (1991). Fungal Burn Wound Infection. Archives of Surgery. 126(1): 44 DOI: 10.1001/archsurg.1991.01410250048008
- Berman J, Krysan DJ (2020). Drug resistance and tolerance in fungi. Nat Rev Microbiol. 18: 319-331 DOI: 10.1038/s41579-019-0322-2
- Clinton A, Carter T (2015). Chronic Wound Biofilms: Pathogenesis and Potential Therapies. Laboratory Medicine. 46(4): 277-284 DOI: 10.1309/LMBNSWKUI4JPN7SO
- Dowd SE, Delton Hanson J, Rees E, Wolcott RD, Zischau AM., et al (2011). Survey of fungi and yeast in polymicrobial infections in chronic wounds. Journal of Wound Care. 20(1): 40-47 DOI: 10.12968/jowc.2011.20.1.40

- Fritsche S, Steinhart H (1999). Occurrence of hormonally active compounds in food: a review. Eur Food Res Technol. 209: 153-179 DOI: 10.1007/s002170050475
- Gerner EW, Meyskens FL (2009). Combination Chemoprevention for Colon Cancer Targeting Polyamine Synthesis and Inflammation. Clinical Cancer Research. 15(3): 758-761 DOI: 10.1158/1078-0432.ccr-08-2235.
- Ghazaryan H, Hovhannisyan A (2022). Comparison of the Pharmacokinetics of Eflornithine after Application of Eflornithine Cream and "Eflornithine: Armenicum" Composition in Rates: In book: Cytotoxicity - Understanding Cellular Damage and Response (Edited by Sukumaran A. and Mahmoud A.M.) 244P, IntechOpen. London, United King. DOI: 10.5772/intechopen.100956, DOI: 10.5772/ intechopen.105742

- Gimeno-LLuch I, Benito-Jardón M, Guerrero-Barberà G, Burday N, Costell M (2022). The Role of the Fibronectin Synergy Site for Skin Wound Healing. Cells. 11(13): 2100 DOI: 10.3390/cells11132100
- Grinnell F, Billingham RE, Burgess L (1981). Distribution of Fibronectin During Wound Healing in Vivo.. Journal of Investigative Dermatology. 76(3): 181-189 DOI: 10.1111/1523-1747.ep12525694
- 11. Hovhannisyan SS, Zilfyan AV, Tarverdyan NA (1987). [Method of modeling an abscess]
 [Published in Russian]. Author's document #1347089 from 221 June 1987
- 12. James GA, Swogger E, Wolcott R, deLancey PE, Secor P., et al (2008). Biofilms in chronic wounds. 16(1): 37-44 DOI: 10.1111/j.1524-475x.2007.00321.x
- Kapoor G, Saigal S, Elongavan A (2017). Action and resistance mechanisms of antibiotics A guide for clinicians, Journal of Anaesthesiology Clinical Pharmacology. 33(3): 300-305 DOI: 10.4103/joacp.JOACP_349_15
- *14. Lenselink EA (2015).* Role of fibronectin in normal wound healing. International Wound Journal. 12(3): 313-316 DOI: 10.1111/iwj.12109
- 15. Meyskens FL, Gerner EW (1999). Development of difluoromethylornithine (DFMO) as a chemoprevention agent. Clin Cancer Res. 5: 945-951
- 16. Michael AJ (2018). Polyamine function in archaea and bacteria. Journal of Biological Chemistry. jbc.TM118.005670–. DOI: 10.1074/jbc.TM118.005670
- 17. Percival SL, McCarty SM, Lipsky B (2015). Biofilms and Wounds: An Overview of the Evidence. Advances in Wound Care. 4(7): 373-381 DOI: 10.1089/wound.2014.0557
- Pfaller MA (2012). Antifungal Drug Resistance: Mechanisms, Epidemiology, and Consequences for Treatment. 125(1S) DOI: 10.1016/j.amjmed.2011.11.001

- Pruskowski KA, Mitchell TA, Kiley JL, Wellington T, Britton GW, Cancio LC (2021). Diagnosis and Management of Invasive Fungal Wound Infections in Burn Patients. Eur Burn J. 2: 168-183 DOI: 10.3390/ebj2040013
- 20. Reygaert WC (2018). An overview of the antimicrobial resistance mechanisms of bacteria. AIMS Microbiology. 4(3): 482-501 DOI: 10.3934/microbiol.2018.3.482
- Rocha RO, Wilson RA (2018). Essential, deadly, enigmatic: Polyamine metabolism and roles in fungal cells. Fungal Biology Reviews. S1749461317300829–. DOI: 10.1016/j. fbr.2018.07.003
- 22. Saha SK, Pathak NN (2021). Fundamentals of Animal Nutrition. Springer. 269p DOI: 10.1007/978-981-15-9125-9
- 23. Shah P, Swiatlo E (2008). A multifaceted role for polyamines in bacterial pathogens. Mol. Microbiol. 68: 4-16 DOI: 10.1111/j.1365-2958.2008.06126.x.
- 24. Valdés-Santiago L, Cervantes-Chávez JA, León-Ramírez CG, Ruiz-Herrera J (2012).
 Polyamine Metabolism in Fungi with Emphasis on Phytopathogenic Species. Journal of Amino Acids. 1-13 DOI: 10.1155/2012/837932
- 25. Valdés-Santiago L, Ruiz-Herrera J (2014). Stress and polyamine metabolism in fungi. Frontiers in Chemistry. 1: DOI: 10.3389/ fchem.2013.00042
- 26. Wallace H, Fraser A (2004). Inhibitors of polyamine metabolism: Review article. Amino Acids. 26: 353-365 DOI: 10.1007/s00726-004-0092-6
- 27. Wallace HM (2009). The polyamines: past, present and future. Essays in Biochemistry. 46: 1-10 DOI: 10.1042/BSE0460001
- 28. Zilfyan AV, Avagyan SA, Ghazaryan AV (2016). The Effect of "Armenicum" paste on the Course of Induced Wound process, Lambert Academic Publishing, Germany

THE NEW ARMENIAN MEDICAL JOURNAL



Volume19 (2025). Issue 1



CONTENTS

- 4. Stilidi E.I., Kliaritskaia I.L., Maksimova E.V., Moshko Yu.A. Chronic hepatitis c with cryoglobulinemia: features and manifestations
- 10. Ćorić N., Banjari I., Rolić T., Marijanović I. NUTRITIONAL AND HEALTH STATUS OF COLORECTAL CANCER PATIENTS - BASELINE STUDY
- 20. Azatyan V.Yu., Yessayan L.K., Poghosyan M.A., Shmavonyan M.V., Sahakyan K.T., Muradyan A.A.

CHARACTERISTICS OF MORPHOLOGICAL ELEMENTS OF LESIONS OF THE ORAL MUCOSA IN PATIENTS WITH HIV INFECTION

31. GHAZARYAN H.V.

THE EFFECT OF THE MEDICINAL COMPOSITION "EFLORNITHINE-ARMENICUM" ON THE PROGRESSION OF THE INFLAMMATORY PROCESS IN AN EXPERIMENTALLY INDUCED AEROBIC WOUND

38. SHAHBAZYAN S.S., TER-AVETIKYAN Z.A., BADALOVA ZH.E.

EVALUATION OF KNOWLEDGE AND ATTITUDE REGARDING MORBID OBESITY AND BARIATRIC SURGERY PRACTICE: AN OBSERVATIONAL ANALYTICAL STUDY IN A NATIONALLY REPRESENTATIVE SAMPLE OF ARMENIAN POPULATION

50. ZILFYAN A.V., AVAGYAN A.S., MURADYAN A.A.

THE ROLE OF RESIDENT BACTERIAL-FUNGAL INTERACTIONS IN BIOFILM FORMATION DURING WOUND INFECTIONS: DOES BIOFILM FORMATION IN ECOLOGICAL NICHES CONTRIBUTE TO NORMAL FUNCTIONING IN VERTEBRATE MAMMALS?

61. Mohammed N.D., Raghavendra R., Arjun B., Aishwarya C., Sujatha B.S.

EFFECTIVENESS OF THERAPEUTIC PLASMA EXCHANGE IN COMPARISON WITH STANDARD OF CARE IN THE TREATMENT OF YELLOW PHOSPHORUS POISONING: AN OBSERVATIONAL STUDY IN SOUTH INDIAN POPULATION

68. Nazaryan L.G., Barseghyan A.B., Simonyan M.H. Consumer behavior in acute diarrhea treatment: analyzing trust in pharmacy employees

75. Aghahosseini F., Omidsalar P., Akhbari P.

THE FIRST REPORT OF GRAPHITE TATTOO IN THE SOFT PALATE: A NOVEL CASE WITH A REVIEW OF ARTICLES

- 81. BARSEGHYAN A.B., DZOAGBE H.Y., GINOVYAN G.G., NAZARYAN L.G., SIMONYAN M.H. ASSESSMENT OF VITAMIN USE AND SELF-MEDICATION PRACTICES AMONG CONSUMERS
- 87. KRISTANTO R., JUNITHA K., SUYANTO H., PHARMAWATI M., YUDIANTO A. SEX DETERMINATION USING CONFOCAL RAMAN MICROSCOPE WITH CHEMOMETRIC METHOD FROM DENTAL SAMPLE AND CONFIRMATION BY AMELOGENIN GENE

95. RAPYAN A.A., CHOPIKYAN A.S., SARGSYAN T.M., SISAKIAN H.S.

COMPARATIVE OUTCOMES FOLLOWING PERCUTANEOUS CORONARY INTERVENTION AND CONSERVATIVE TREATMENT IN ELDERLY PATIENTS WITH ACUTE MYOCARDIAL INFARCTION: SINGLE CENTER RETROSPECTIVE COHORT ANALYSIS

104. Restrepo Gil E., Aguirre Correa L.A., Cardona Maya W.D.

KNOWLEDGE AND PERCEPTIONS ABOUT THE DIGITAL RECTAL EXAMINATION: EXPERIENCES IN COLOMBIA

112. Gekhaev A.U., Isakova F.S., Gadaev I.Sh.

ROLE OF CORTISOL IN THE CARCINOGENESIS OF LARYNGEAL CANCER

THE NEW ARMENIAN MEDICAL JOURNAL

Volume19 (2025). Issue 1





The Journal is founded by Yerevan State Medical University after M. Heratsi.

Rector of YSMU

Armen A. Muradyan

Address for correspondence:

Yerevan State Medical University 2 Koryun Street, Yerevan 0025, Republic of Armenia

Phones:

(+37410) 582532 YSMU (+37493 588697 Editor-in-Chief **Fax:** (+37410) 582532 **E-mail:**namj.ysmu@gmail.com, ysmiu@mail.ru **URL:**http//www.ysmu.am

Our journal is registered in the databases of Scopus, EBSCO and Thomson Reuters (in the registration process)



Scopus EBSC Reuters

Copy editor: Tatevik R. Movsisyan

LLC Print in "Monoprint" LLC

Director: Armen Armenaakyan Andraniks St., 96/8 Bulding Yerevan, 0064, Armenia Phone: (+37491) 40 25 86 E-mail: monoprint1@mail.ru

Editor-in-Chief

Arto V. Zilfyan (Yerevan, Armenia)

Deputy Editors

Hovhannes M. **Manvelyan** (Yerevan, Armenia) Hamayak S. **Sisakyan** (Yerevan, Armenia)

Executive Secretary

Stepan A. Avagyan (Yerevan, Armenia)

Editorial Board

Armen A. **Muradyan** (Yerevan, Armenia) Drastamat N. **Khudaverdyan** (Yerevan, Armenia) Levon M. **Mkrtchyan** (Yerevan, Armenia)

Foregin Members of the Editorial Board

Carsten N. GUTT (Memmingen, Germay) Muhammad MIFTAHUSSURUR (Indonesia) Alexander WOODMAN (Dharhan, Saudi Arabia)

Coordinating Editor (for this number)

Hesam Adin Atashi (Tehran, Iran)

Editorial Advisory Council

Mahdi Esmaeilzadeh (Mashhad, Iran) Ara S. Babloyan (Yerevan, Armenia) Aram Chobanian (Boston, USA) Luciana **Dini** (Lecce, Italy) Azat A. Engibaryan (Yerevan, Armenia) Ruben V. Fanarjyan (Yerevan, Armenia) Gerasimos Filippatos (Athens, Greece) Gabriele Fragasso (Milan, Italy) Samvel G. Galstyan (Yerevan, Armenia) Arthur A. Grigorian (Macon, Georgia, USA) Armen Dz. Hambardzumyan (Yerevan, Armenia) Seyran P. Kocharyan (Yerevan, Armenia) Aleksandr S. Malayan (Yerevan, Armenia) Mikhail Z. Narimanyan (Yerevan, Armenia) Levon N. Nazarian (Philadelphia, USA) Yumei Niu (Harbin, China) Linda F. Noble-Haeusslein (San Francisco, USA) Arthur K. Shukuryan (Yerevan, Armenia) Suren A. Stepanyan (Yerevan, Armenia) Gevorg N. Tamamyan (Yerevan, Armenia) Hakob V. Topchyan (Yerevan, Armenia) Alexander Tsiskaridze (Tbilisi, Georgia) Konstantin B. Yenkoyan (Yerevan, Armenia) Peijun Wang (Harbin, Chine)